KPL Protein A Agarose



<u>Size</u>	
5	mL

DESCRIPTION

5710-0005 (223-50-01)

Item No.

KPL Protein A Agarose consists of native protein A immobilized onto 4% cross linked agarose beads. It is designed specifically for the binding of immunoglobulins for both laboratory and process scale applications. The protein A molecule is very heat stable and retains its native conformation even after exposure to denaturing reagents such as 4M urea, 4M guanidine thiocyanate or 6M guanidine hydrochloride⁽¹⁾. Protein A binds specifically to the Fc region of immunoglobulin molecules of many mammalian species without disturbing their binding of antigen.

Covalently coupled Protein A Agarose has been extensively used for the isolation of a wide variety of immunoglobulin molecules from several mammalian species. Table 1 describes the relative affinity of immobilized Protein A for different antibody species and subclasses.

FORM/STORAGE

KPL Protein A Agarose is supplied in a volume of 7 mL consisting of 5 mL KPL Protein A Agarose suspended in 20% ethanol/PBS. Store refrigerated at 2-8°C. Stable for a minimum of 1 year from date of receipt when stored at 2-8°C. Non-sterile.

SPECIFICATIONS

Ligand density:	~ 6mg Protein A/mL gel
Bead structure: agarose	4% cross-linked
Bead size range:	45 - 165 µm
Recommended working pH:	3 9
Binding capacity:	>35mg/mL Human IgG

Note: Different immunoglobulins derived from the same species and from the same subclass can demonstrate deviations in the binding capacity.

Table 1. Relative Affinity of Immobilized Protein Afor Various Antibody Species and Subclasses ofpolyclonal andEC(2)

	es/Subclass	Protein A
	OCLONAL	
Humar		
	lgG ₁	++++
	IgG 2	++++
	lgG 3	
	lgG 4	++++
Mouse		
	lgG 1	+
	IgG 2a	++++
	IgG 2b	+++
Det	lgG 3	++
Rat		
	IgG 1	
	IgG 2a	
	IgG 2b	
	lgG 2c	+
POLY	CLONAL	
Rabbit		++++
Cow		++
Horse		++
Goat		-
Guinea	a pig	++++
Sheep		+/-
Pig		+++
Rat		+/-
Mouse)	++
Chicke		
Humar		++++
Humar		
Humar		
Humar	n IgA	

--- (weak or no binding) \rightarrow ++++ (Strong binding)

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PROCEDURE

PURIFICATION OF IgG MOLECULES

1. User Supplied Materials

- a. Buffers: see Section 2 below.
- Disposable column with frits and reusable caps. SeraCare recommends Pharmacia Biotech PD-10 empty disposable columns or equivalent.

2. Buffer Preparation

- a. Wash/Binding Buffer: KPL Wash/Binding Buffer or prepare 0.1 M Sodium phosphate, 0.15 M NaCl, pH 7.4.
- b. Elution Buffer: KPL Elution Buffer or prepare 0.2 M Glycine, pH 3.0 ±0.15.
 Storage Buffer: KPL Storage Buffer or prepare 0.01 M NaH₂PO₄, 0.15M NaCl, 2.7 mM KCl, pH 7.4,20% ethanol
- 3. Sample Preparation: To insure proper ionic strength and pH are maintained for optimal binding, it is necessary to dilute serum samples, ascites fluid or tissue culture supernatant at least 1/1 with binding buffer. Alternatively, the sample may be dialyzed overnight against wash/binding buffer. SeraCare recommends using a 12,000 MW cutoff dialysis tubing with at least 2 buffer exchanges. Remove any particulate matter from the sample by centrifugation or filtration through a 0.8 µm filter.

4. Column and Resin Preparation:

- a. Pour 20% Ethanol in the bottom of a petri dish or in a flat bottomed container. Float the frit on top of the ethanol. Using the large round end of a 1 mL pipette tip, press the frit firmly into the ethanol to force air out. Repeat this step until the frit is completely wet.
- b. Push the frit into the barrel of the column until it rests firmly on the bottom.
- c. With the cap removed, clip the end of the column to create a hole to allow liquid to flow through.
- d. Wash the frit with 5 column volumes of 1X KPL Wash/Binding Buffer.
- e. Prepare a 1/1 suspension of resin in 1X Wash/Binding buffer. The required amount of agarose per mg of immunoglobulin being purified can be estimated by the binding capacity.

Recommended Column Volumes:

Antibody Source	Recommended bed volume (mL) per mL sample
Immune Serum	2 mL
Tissue Culture Supernatant (with 10% fetal bovine serum)	0.2 mL
Tissue Culture Supernatant (serum-free)	0.01 mL
Ascites Fluid	2 mL

f. Pour slurry into column. Allow column to flow by gravity to pack the column bed.

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S	ize
5	mL

- 7. **Clean-in-Place:** With certain applications, substances which contain denatured proteins or lipids do not elute in the regeneration procedure. The following steps can be taken to clean the column:
 - a. To remove strongly bound hydrophobic proteins, lipoproteins and lipids, wash the column with a non-ionic detergent (e.g. 0.1% Triton X-100) at 37°C, with a contact time of ~1 minute.
 - Immediately re-equilibrate the column with
 5 10 CV of 1X KPL Wash/Binding Buffer.
 - c. As an alternative, wash the column with 70% ethanol. Allow the column to stand for 12 hours.
 - d. Re-equilibrate the column with 5 10 CV of 1X KPL Wash/Binding Buffer.
 - e. To remove precipitated or denatured substances, wash the column with 2 CV of 6M guanidine hydrochloride. Immediately reequilibrate the column with 5 - 10 CV of 1X KPL Wash/Binding Buffer (see step 6).
- 8. **Resin Storage:** Store affinity matrix in storage buffer at 2-8°C. Do not store the matrix frozen or at room temperature. The matrix can be stored in the column by sealing the outlets or remove from the column and stored as a slurry.

IMMUNOPRECIPITATION

For immunoprecipitation protocols, see references 3 - 5.

PRODUCT SAFETY AND HANDLING

See SDS (Safety Data Sheet) for this product.

REFERENCES

1. Surolia, A., Pain, D. and Khan, M.I., (1982). *Trends Biochem. Sci.*, 7, 74 - 76.

2. Harlow, E. and Lane, D. eds. (1988). Antibodies, A Laboratory Manual. Cold Spring Harbor Laboratory, N.Y., 617 - 618.

3. Langone, J.J, (1982). *J. Immunological Methods*, 55, 277 - 296.

4. Lindmark, R., Thoren-Tolling, K., Sjoquist, J., (1983). J. Immunological Methods, 62, 1 - 13.

5. Thurston, C.F. and Henley, L.F., (1988). *in* Walker, J.M., ed. Methods in Molecular Biology, Vol. 3- New Protein Techniques. Humana Press: Clifton, N.J., 149 - 158.

RELATED PRODUCTS CAT. NO.

KPL Protein A Agarose Kit	5710-0009 (553-50-01)
KPL Protein G Agarose Kit	5720-0004 (553-51-00)
KPL Protein G Agarose	5720-0002 (223-51-01)

The product listed herein is for research use only and is not intended for use in human or clinical diagnosis.